Purpose: To try Ayanthi imbalance dNTPs in transformation whether we get better ✱.

L 1	=	Taq 1 U	(200 µM each nucleotide)	1
L 4	=	Taq 4 U	" "	2
Taq: H 1	=	" 1 U	200 µM dA & rest 1 mM	3
H 4	=	" 4 U	" "	4

LD 1	=	Taq + DV	1	(200)
LD 4	=	"	4	"
HD 1	=	"	1	(200 + 1 × 3)
HD 4	=	"	4	"

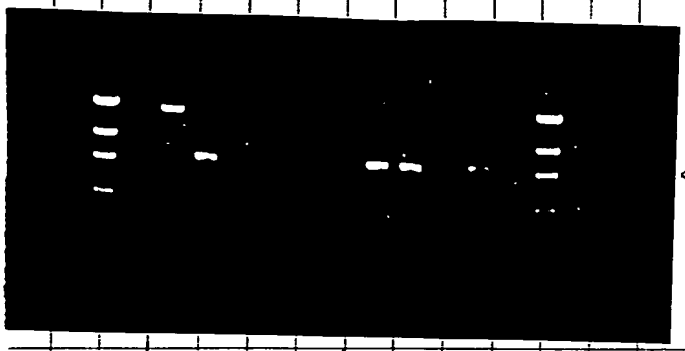
- Duplicates of each reactions pooled together, ethanol pptd after a phenol chloroform extraction.
- Resuspended in 15 µl ~~Reaction~~ TE cut with Afl III and Aat II in NEB buffer overnight at 37°.
- 2 µl of each run on gel to see the digestion is comp
- Even though Ayanthi said there is enough product in PCR r some of them didn't show up on the gel after all the purification steps.
- Since there is not much time to gel purify the fragments whole reaction as such, was used in the ligation reaction ~~2~~ 10 µl of it.

Witnessed & Understood by me, [signature] Date 12/1?/94

Invented by _______ Date 12/15/94

Recorded by [signature]

ag No.__



Vector | Tag Rx | Tag + Dx

ligation

1.	Vector	only
2.	Vector	+ rest
3.	Tag	1 1
4		1 4
5		4 1
6		4 4
7	T+Dx	1 D 1
8		1 D 4
9		4 D 1
10		4 D 4

10 pul of Restriction dont know exactly how much insert is there

Vector	1 pul
insert	10 pul
Ligase	1 pul
10 5x buffer	4 pul
	20 pul at 25°, 3 hrs.

Transformed all 10 (1 - 10 above) & Control insert movement pre.

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sed & Understood by me,	Date 12/16/94	Invented by	Date
		Recorded by K. Shaaman	12/16/94

[redacted]